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Munich, 19th June 2001

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Faust

New nucleotide sequences which code for the citA gene

The invention provides nucleotide sequences from coryneform bacteria which code for the citA gene and a process for the fermentative preparation of amino acids, in particular L-lysine, by attenuation of the citA gene. The citA gene codes for the sensor kinase Cit A of a two-component system.

Prior art

L-Amino acids, in particular L-lysine, are used in human medicine and in the pharmaceuticals industry, in the foodstuffs industry and very particularly in animal nutrition.

It is known that amino acids are prepared by fermentation from strains of coryneform bacteria, in particular *Corynebacterium glutamicum*. Because of their great importance, work is constantly being undertaken to improve the preparation processes. Improvements to the process can relate to fermentation measures, such as, for example, stirring and supply of oxygen, or the composition of the nutrient media, such as, for example, the sugar concentration during the fermentation, or the working up to the product form by, for example, ion exchange chromatography, or the intrinsic output properties of the microorganism itself.

Methods of mutagenesis, selection and mutant selection are used to improve the output properties of these microorganisms. Strains which are resistant to antimetabolites or are auxotrophic for metabolites of regulatory importance and which produce amino acids are obtained in this manner.

Methods of the recombinant DNA technique have also been employed for some years for improving the strain of *Corynebacterium* strains which produce L-amino acids.

Object of the invention

The inventors had the object of providing new measures for improved fermentative preparation of amino acids, in particular L-lysine.

5 Description of the invention

If L-lysine or lysine are mentioned in the following, this also means the salts, such as e. g. lysine monohydrochloride or lysine sulfate.

The invention provides an isolated polynucleotide from
10 coryneform bacteria, comprising a polynucleotide sequence which codes for the *citA* gene chosen from the group consisting of

- a) polynucleotide which is identical to the extent of at least 70 % to a polynucleotide which codes for a
15 polypeptide which comprises the amino acid sequence of SEQ ID No. 2,
- b) polynucleotide which codes for a polypeptide which comprises an amino acid sequence which is identical to the extent of at least 70 % to the amino acid sequence
20 of SEQ ID No. 2,
- c) polynucleotide which is complementary to the polynucleotides of a) or b), and
- d) polynucleotide comprising at least 15 successive nucleotides of the polynucleotide sequence of a), b)
25 or c),

the polypeptide preferably having the activity of sensor kinase CitA.

The invention also provides the abovementioned polynucleotide, this preferably being a DNA which is
30 capable of replication, comprising:

- (i) the nucleotide sequence shown in SEQ ID No.1 or
- (ii) at least one sequence which corresponds to sequence (i) within the range of the degeneration of the genetic code, or
- 5 (iii) at least one sequence which hybridizes with the sequences complementary to sequences (i) or (ii), and optionally
- (iv) sense mutations of neutral function in (i).

The invention also provides:

10 a polynucleotide, in particular DNA, which is capable of replication and comprises the nucleotide sequence as shown in SEQ ID No.1;

a polynucleotide which codes for a polypeptide which comprises the amino acid sequence as shown in SEQ ID
15 No. 2;

a vector containing parts of the polynucleotide according to the invention, but at least 15 successive nucleotides of the sequence claimed

and coryneform bacteria in which the *citA* gene is
20 attenuated, in particular by an insertion or deletion.

The invention also provides polynucleotides which substantially comprise a polynucleotide sequence, which are obtainable by screening by means of hybridization of a corresponding gene library of a coryneform bacterium, which
25 comprises the complete gene or parts thereof, with a probe which comprises the sequence of the polynucleotide according to the invention or a fragment thereof, and isolation of the polynucleotide sequence mentioned.

Polynucleotides comprising the sequences according to the
30 invention are suitable as hybridization probes for RNA,

cDNA and DNA, in order to isolate, in the full length, nucleic acids or polynucleotides or genes which code for CitA protein or to isolate those nucleic acids or polynucleotides or genes which have a high similarity with
5 the sequence of the citA gene.

Polynucleotides comprising the sequences according to the invention are furthermore suitable as primers with the aid of which DNA of genes which code for the CitA protein can be prepared by the polymerase chain reaction (PCR).

10 Such oligonucleotides which serve as probes or primers comprise at least 30, preferably at least 20, very particularly preferably at least 15 successive nucleotides. Oligonucleotides which have a length of at least 40 or 50 nucleotides are also suitable.

15 "Isolated" means separated out of its natural environment.

"Polynucleotide" in general relates to polyribonucleotides and polydeoxyribonucleotides, it being possible for these to be non-modified RNA or DNA or modified RNA or DNA.

The polynucleotides according to the invention include a
20 polynucleotide according to SEQ ID No. 1 or a fragment prepared therefrom and also those which are at least 70 %, preferably at least 80 % and in particular at least 90 % to 95 % identical to the polynucleotide according to SEQ ID No. 1 or a fragment prepared therefrom.

25 "Polypeptides" are understood as meaning peptides or proteins which comprise two or more amino acids bonded via peptide bonds.

The polypeptides according to the invention include a polypeptide according to SEQ ID No. 2, in particular those
30 with the biological activity of the CitA protein and also those which are at least 70 %, preferably at least 80 % and in particular at least 90 % to 95 % identical to the

polypeptide according to SEQ ID No. 2 and have the activity mentioned.

The invention moreover relates to a process for the fermentative preparation of amino acids, in particular L-lysine, using coryneform bacteria which in particular already produce amino acids and in which the nucleotide sequences which code for the *citA* gene are attenuated, in particular eliminated or expressed at a low level.

The term "attenuation" in this connection describes the reduction or elimination of the intracellular activity of one or more enzymes (proteins) in a microorganism which are coded by the corresponding DNA, for example by using a weak promoter or using a gene or allele which codes for a corresponding enzyme with a low activity or inactivates the corresponding gene or enzyme (protein), and optionally combining these measures.

The microorganisms which the present invention provides can prepare amino acids, in particular L-lysine, from glucose, sucrose, lactose, fructose, maltose, molasses, starch, cellulose or from glycerol and ethanol. They can be representatives of coryneform bacteria, in particular of the genus *Corynebacterium*. Of the genus *Corynebacterium*, there may be mentioned in particular the species *Corynebacterium glutamicum*, which is known among experts for its ability to produce L-amino acids.

Suitable strains of the genus *Corynebacterium*, in particular of the species *Corynebacterium glutamicum* (*C. glutamicum*), are in particular the known wild-type strains

Corynebacterium glutamicum ATCC13032
Corynebacterium acetoglutamicum ATCC15806
Corynebacterium acetoacidophilum ATCC13870
Corynebacterium melassecola ATCC17965
Corynebacterium thermoaminogenes FERM BP-1539
Brevibacterium flavum ATCC14067

Brevibacterium lactofermentum ATCC13869 and
Brevibacterium divaricatum ATCC14020

or L-amino acid-producing mutants or strains prepared
therefrom, such as, for example, the L-lysine-producing
5 strains

Corynebacterium glutamicum FERM-P 1709
Brevibacterium flavum FERM-P 1708
Brevibacterium lactofermentum FERM-P 1712
Corynebacterium glutamicum FERM-P 6463
10 *Corynebacterium glutamicum* FERM-P 6464
Corynebacterium glutamicum DM58-1
Corynebacterium glutamicum DG52-5
Corynebacterium glutamicum DSM 5715 and
Corynebacterium glutamicum DSM 12866

15 The inventors have succeeded in isolating the new *citA* gene
of *C. glutamicum* which codes for the Cita protein and which
is a sensor kinase of a two-component system.

To isolate the *citA* gene or also other genes of *C.*
glutamicum, a gene library of this microorganism is first
20 set up in *Escherichia coli* (*E. coli*). The setting up of
gene libraries is described in generally known textbooks
and handbooks. The textbook by Winnacker: *Gene und Klone, Eine Einführung in die Gentechnologie* [Genes and Clones, An
Introduction to Genetic Engineering] (Verlag Chemie,
25 Weinheim, Germany, 1990), or the handbook by Sambrook et
al.: *Molecular Cloning, A Laboratory Manual* (Cold Spring
Harbor Laboratory Press, 1989) may be mentioned as an
example. A well-known gene library is that of the *E. coli*
K-12 strain W3110 set up in λ -vectors by Kohara et al.
30 (Cell 50, 495-508 (1987)). Bathe et al. (Molecular and
General Genetics, 252:255-265, 1996) describe a gene
library of *C. glutamicum* ATCC13032, which was set up with
the aid of the cosmid vector SuperCos I (Wahl et al., 1987,
Proceedings of the National Academy of Sciences USA,

84:2160-2164) in the E. coli K-12 strain NM554 (Raleigh et al., 1988, Nucleic Acids Research 16:1563-1575). Börmann et al. (Molecular Microbiology 6(3), 317-326 (1992)) in turn describe a gene library of C. glutamicum ATCC13032
5 using the cosmid pHC79 (Hohn and Collins, 1980, Gene 11, 291-298).

To prepare a gene library of C. glutamicum in E. coli it is also possible to use plasmids such as pBR322 (Bolivar, 1979, Life Sciences, 25, 807-818) or pUC9 (Vieira et al.,
10 1982, Gene, 19:259-268). Suitable hosts are, in particular, those E. coli strains which are restriction- and recombination-defective, such as, for example, the strain DH5 α (Jeffrey H. Miller: "A Short Course in Bacterial Genetics, A Laboratory Manual and Handbook for Escherichia
15 coli and Related Bacteria", Cold Spring Harbor Laboratory Press, 1992).

The long DNA fragments cloned with the aid of cosmids or other λ -vectors can then be subcloned in turn into the usual vectors suitable for DNA sequencing.

20 Methods of DNA sequencing are described, inter alia, by Sanger et al. (Proceedings of the National Academy of Sciences of the United States of America USA, 74:5463-5467, 1977).

The resulting DNA sequences can then be investigated with
25 known algorithms or sequence analysis programs, such as e.g. that of Staden (Nucleic Acids Research 14, 217-232(1986)), that of Marck (Nucleic Acids Research 16, 1829-1836 (1988)) or the GCG program of Butler (Methods of Biochemical Analysis 39, 74-97 (1998)).

30 The new DNA sequence of C. glutamicum which codes for the *citA* gene and which, as SEQ ID No. 1, is a constituent of the present invention has been found in this way. The amino acid sequence of the corresponding protein has furthermore been derived from the present DNA sequence by the methods

described above. The resulting amino acid sequence of the citA gene product is shown in SEQ ID No. 2.

Coding DNA sequences which result from SEQ ID No. 1 by the degeneracy of the genetic code are also a constituent of
5 the invention. In the same way, DNA sequences which hybridize with SEQ ID No. 1 or parts of SEQ ID No. 1 are a constituent of the invention. Conservative amino acid exchanges, such as e.g. exchange of glycine for alanine or of aspartic acid for glutamic acid in proteins, are
10 furthermore known among experts as "sense mutations" which do not lead to a fundamental change in the activity of the protein, i.e. are of neutral function. It is furthermore known that changes on the N and/or C terminus of a protein cannot substantially impair or can even stabilize the
15 function thereof. Information in this context can be found by the expert, inter alia, in Ben-Bassat et al. (Journal of Bacteriology 169:751-757 (1987)), in O'Regan et al. (Gene 77:237-251 (1989)), in Sahin-Toth et al. (Protein Sciences 3:240-247 (1994)), in Hochuli et al. (Bio/Technology
20 6:1321-1325 (1988)) and in known textbooks of genetics and molecular biology. Amino acid sequences which result in a corresponding manner from SEQ ID No. 2 are also a constituent of the invention.

Finally, DNA sequences which are prepared by the polymerase
25 chain reaction (PCR) using primers which result from SEQ ID No. 1 are a constituent of the invention. Such oligonucleotides typically have a length of at least 15 nucleotides.

Instructions for identifying DNA sequences by means of
30 hybridization can be found by the expert, inter alia, in the handbook "The DIG System Users Guide for Filter Hybridization" from Boehringer Mannheim GmbH (Mannheim, Germany, 1993) and in Liebl et al. (International Journal of Systematic Bacteriology 41: 255-260 (1991)). The
35 hybridization takes place under stringent conditions, that

is to say only hybrids in which the probe and target sequence, i. e. the polynucleotides treated with the probe, are at least 70 % identical are formed. It is known that the stringency of the hybridization, including the washing
5 steps, is influenced or determined by varying the buffer composition, the temperature and the salt concentration. The hybridization reaction is preferably carried out under a relatively low stringency compared with the washing steps (Hybaid Hybridisation Guide, Hybaid Limited, Teddington,
10 UK, 1996). A 5x SSC buffer at a temperature of approx. 50 - 68°C, for example, can be employed for the hybridization reaction. Probes can also hybridize here with polynucleotides which are less than 70 % identical to the sequence of the probe. Such hybrids are less stable and are
15 removed by washing under stringent conditions. This can be achieved, for example, by lowering the salt concentration to 2x SSC and optionally subsequently 0.5x SSC (The DIG System User's Guide for Filter Hybridisation, Boehringer Mannheim, Mannheim, Germany, 1995) a temperature of approx.
20 50 - 68°C being established. Polynucleotide fragments which are, for example, at least 70 % or at least 80 % or at least 90 % to 95 % identical to the sequence of the probe employed can be isolated by increasing the hybridization temperature stepwise from 50 to 68°C in steps of approx. 1
25 - 2°C. Further instructions on hybridization are obtainable on the market in the form of so-called kits (e.g. DIG Easy Hyb from Roche Diagnostics GmbH, Mannheim, Germany, Catalogue No. 1603558).

Instructions for amplification of DNA sequences with the
30 aid of the polymerase chain reaction (PCR) can be found by the expert, inter alia, in the handbook by Gait: Oligonukleotides: a Practical Approach (IRL Press, Oxford, UK, 1984) and in Newton and Graham: PCR (Spektrum Akademischer Verlag, Heidelberg, Germany, 1994).

In the work on the present invention, it has been found that coryneform bacteria produce amino acids, in particular L-lysine, in an improved manner after attenuation of the *citA* gene.

- 5 To achieve an attenuation, either the expression of the *citA* gene or the catalytic properties of the enzyme protein can be reduced or eliminated. The two measures can optionally be combined.

The reduction in gene expression can take place by suitable
10 culturing or by genetic modification (mutation) of the signal structures of gene expression. Signal structures of gene expression are, for example, repressor genes, activator genes, operators, promoters, attenuators, ribosome binding sites, the start codon and terminators.
15 The expert can find information on this e.g. in the patent application WO 96/15246, in Boyd and Murphy (Journal of Bacteriology 170: 5949 (1988)), in Voskuil and Chambliss (Nucleic Acids Research 26: 3548 (1998)), in Jensen and Hammer (Biotechnology and Bioengineering 58: 191 (1998)),
20 in Pátek et al. (Microbiology 142: 1297 (1996)), Vasicova et al. (Journal of Bacteriology 181: 6188 (1999)) and in known textbooks of genetics and molecular biology, such as e.g. the textbook by Knippers ("Molekulare Genetik [Molecular Genetics]", 6th edition, Georg Thieme Verlag, Stuttgart, Germany, 1995) or that by Winnacker ("Gene und
25 Klone [Genes and Clones]", VCH Verlagsgesellschaft, Weinheim, Germany, 1990).

Mutations which lead to a change or reduction in the catalytic properties of enzyme proteins are known from the
30 prior art; examples which may be mentioned are the works by Qiu and Goodman (Journal of Biological Chemistry 272: 8611-8617 (1997)), Sugimoto et al. (Bioscience Biotechnology and Biochemistry 61: 1760-1762 (1997)) and Möckel ("Die Threonindehydratase aus *Corynebacterium glutamicum*:
35 Aufhebung der allosterischen Regulation und Struktur des

Enzymes [Threonine dehydratase from *Corynebacterium glutamicum*: Cancelling the allosteric regulation and structure of the enzyme]", Reports from the Jülich Research Centre, Jül-2906, ISSN09442952, Jülich, Germany, 1994).

- 5 Summarizing descriptions can be found in known textbooks of genetics and molecular biology, such as e.g. that by Hagemann ("Allgemeine Genetik [General Genetics]", Gustav Fischer Verlag, Stuttgart, 1986).

- Possible mutations are transitions, transversions,
10 insertions and deletions. Depending on the effect of the amino acid exchange on the enzyme activity, missense mutations or nonsense mutations are referred to. Insertions or deletions of at least one base pair (bp) in a gene lead to frame shift mutations, as a consequence of which
15 incorrect amino acids are incorporated or translation is interrupted prematurely. Deletions of several codons typically lead to a complete loss of the enzyme activity. Instructions on generation of such mutations are prior art and can be found in known textbooks of genetics and
20 molecular biology, such as e.g. the textbook by Knippers ("Molekulare Genetik [Molecular Genetics]", 6th edition, Georg Thieme Verlag, Stuttgart, Germany, 1995), that by Winnacker ("Gene und Klone [Genes and Clones]", VCH Verlagsgesellschaft, Weinheim, Germany, 1990) or that by
25 Hagemann ("Allgemeine Genetik [General Genetics]", Gustav Fischer Verlag, Stuttgart, 1986).

A common method of mutating genes of *C. glutamicum* is the method of gene disruption and gene replacement described by Schwarzer and Pühler (Bio/Technology 9, 84-87 (1991)).

- 30 In the method of gene disruption a central part of the coding region of the gene of interest is cloned in a plasmid vector which can replicate in a host (typically *E. coli*), but not in *C. glutamicum*. Possible vectors are, for example, pSUP301 (Simon et al., Bio/Technology 1, 784-791
35 (1983)), pK18mob or pK19mob (Schäfer et al., Gene 145, 69-

73 (1994)), pK18mobsacB or pK19mobsacB (Jäger et al.,
Journal of Bacteriology 174: 5462-65 (1992)), pGEM-T
(Promega corporation, Madison, WI, USA), pCR2.1-TOPO
(Shuman (1994). Journal of Biological Chemistry 269:32678-
5 84; US Patent 5,487,993), pCR®Blunt (Invitrogen,
Groningen, Holland; Bernard et al., Journal of Molecular
Biology, 234: 534-541 (1993)) or pEM1 (Schrumpf et al,
1991, Journal of Bacteriology 173:4510-4516). The plasmid
vector which contains the central part of the coding region
10 of the gene is then transferred into the desired strain of
C. glutamicum by conjugation or transformation. The method
of conjugation is described, for example, by Schäfer et al.
(Applied and Environmental Microbiology 60, 756-759
(1994)). Methods for transformation are described, for
15 example, by Thierbach et al. (Applied Microbiology and
Biotechnology 29, 356-362 (1988)), Dunican and Shivnan
(Bio/Technology 7, 1067-1070 (1989)) and Tauch et al. (FEMS
Microbiological Letters 123, 343-347 (1994)). After
homologous recombination by means of a "cross-over" event,
20 the coding region of the gene in question is interrupted by
the vector sequence and two incomplete alleles are
obtained, one lacking the 3' end and one lacking the 5'
end. This method has been used, for example, by Fitzpatrick
et al. (Applied Microbiology and Biotechnology 42, 575-580
25 (1994)) to eliminate the recA gene of C. glutamicum.

In the method of gene replacement, a mutation, such as e.g.
a deletion, insertion or base exchange, is established in
vitro in the gene of interest. The allele prepared is in
turn cloned in a vector which is not replicative for C.
30 glutamicum and this is then transferred into the desired
host of C. glutamicum by transformation or conjugation.
After homologous recombination by means of a first "cross-
over" event which effects integration and a suitable second
"cross-over" event which effects excision in the target
35 gene or in the target sequence, the incorporation of the
mutation or of the allele is achieved. This method was

used, for example, by Peters-Wendisch et al. (Microbiology 144, 915 - 927 (1998)) to eliminate the *pyc* gene of *C. glutamicum* by a deletion.

A deletion, insertion or a base exchange can be
5 incorporated into the *citA* gene in this manner.

In addition, it may be advantageous for the production of L-amino acids, in particular L-lysine, to enhance, in particular to over-express, one or more enzymes of the particular biosynthesis pathway, of glycolysis, of
10 anaplerosis, of the pentose phosphate cycle or of amino acid export and optionally regulatory proteins, in addition to attenuation of the *citA* gene.

The term "enhancement" in this connection describes the increase in the intracellular activity of one or more
15 enzymes (proteins) in a microorganism which are coded by the corresponding DNA, for example by increasing the number of copies of the gene or genes, using a potent promoter or using a gene or allele which codes for a corresponding enzyme (protein) having a high activity, and optionally
20 combining these measures.

Thus, for example, for the preparation of L-lysine, at the same time one or more of the genes chosen from the group consisting of

- the *dapA* gene which codes for dihydrodipicolinate
25 synthase (EP-B 0 197 335),
- the *gap* gene which codes for glyceraldehyde 3-phosphate dehydrogenase (Eikmanns (1992), Journal of Bacteriology 174:6076-6086),
- the *zwf* gene which codes for glucose 6-phosphate
30 dehydrogenase (JP-A-09224661),
- the *pyc* gene which codes for pyruvate carboxylase (DE-A-198 31 609),

- the lysE gene which codes for lysine export
(DE-A-195 48 222)
- the lysC gene which codes for a feed back resistant
aspartate kinase (EP-B-0387527; EP-A-0699759; WO
5 00/63388), or
- the zwal gene which codes for the Zwal protein
(DE: 199 59 328.0, DSM 13115)

can be enhanced, in particular over-expressed.

It may furthermore be advantageous for the production of
10 amino acids, in particular L-lysine, in addition to the
attenuation of the citA gene, at the same time for one or
more of the genes chosen from the group consisting of

- the pck gene which codes for phosphoenol pyruvate
carboxykinase (DE 199 50 409.1, DSM 13047),
- 15 • the pgi gene which codes for glucose 6-phosphate
isomerase (US 09/396,478, DSM 12969),
- the poxB gene which codes for pyruvate oxidase
(DE:1995 1975.7, DSM 13114)
- the zwa2 gene which codes for the Zwa2 protein
20 (DE: 199 59,327.2, DSM 13113)

to be attenuated.

In addition to attenuation of the citA gene it may
furthermore be advantageous, for the production of amino
acids, in particular L-lysine, to eliminate undesirable
25 side reactions, (Nakayama: "Breeding of Amino Acid
Producing Microorganisms", in: Overproduction of Microbial
Products, Krumphanzl, Sikyta, Vanek (eds.), Academic Press,
London, UK, 1982).

- The invention also provides the microorganisms prepared according to the invention, and these can be cultured continuously or discontinuously in the batch process (batch culture) or in the fed batch (feed process) or repeated fed
5 batch process (repetitive feed process) for the purpose of production of L-amino acids, in particular L-lysine. A summary of known culture methods is described in the textbook by Chmiel (Bioprozesstechnik 1. Einführung in die Bioverfahrenstechnik [Bioprocess Technology 1. Introduction
10 to Bioprocess Technology] (Gustav Fischer Verlag, Stuttgart, 1991)) or in the textbook by Storhas (Bioreaktoren und periphere Einrichtungen [Bioreactors and Peripheral Equipment] (Vieweg Verlag, Braunschweig/Wiesbaden, 1994)).
- 15 The culture medium to be used must meet the requirements of the particular strains in a suitable manner. Descriptions of culture media for various microorganisms are contained in the handbook "Manual of Methods for General Bacteriology" of the American Society for Bacteriology
20 (Washington D.C., USA, 1981). Sugars and carbohydrates, such as e.g. glucose, sucrose, lactose, fructose, maltose, molasses, starch and cellulose, oils and fats, such as, for example, soya oil, sunflower oil, groundnut oil and coconut fat, fatty acids, such as, for example, palmitic acid,
25 stearic acid and linoleic acid, alcohols, such as, for example, glycerol and ethanol, and organic acids, such as, for example, acetic acid, can be used as the source of carbon. These substances can be used individually or as a mixture.
- 30 Organic nitrogen-containing compounds, such as peptones, yeast extract, meat extract, malt extract, corn steep liquor, soya bean flour and urea, or inorganic compounds, such as ammonium sulfate, ammonium chloride, ammonium phosphate, ammonium carbonate and ammonium nitrate, can be
35 used as the source of nitrogen. The sources of nitrogen can be used individually or as a mixture.

Phosphoric acid, potassium dihydrogen phosphate or dipotassium hydrogen phosphate or the corresponding sodium-containing salts can be used as the source of phosphorus. The culture medium must furthermore comprise salts of
5 metals, such as, for example, magnesium sulfate or iron sulfate, which are necessary for growth. Finally, essential growth substances, such as amino acids and vitamins, can be employed in addition to the abovementioned substances. Suitable precursors can moreover be added to the culture
10 medium. The starting substances mentioned can be added to the culture in the form of a single batch, or can be fed in during the culture in a suitable manner.

Basic compounds, such as sodium hydroxide, potassium hydroxide, ammonia or aqueous ammonia, or acid compounds,
15 such as phosphoric acid or sulfuric acid, can be employed in a suitable manner to control the pH of the culture. Antifoams, such as, for example, fatty acid polyglycol esters, can be employed to control the development of foam. Suitable substances having a selective action, such as, for
20 example, antibiotics, can be added to the medium to maintain the stability of plasmids. To maintain aerobic conditions, oxygen or oxygen-containing gas mixtures, such as, for example, air, are introduced into the culture. The temperature of the culture is usually 20°C to 45°C, and
25 preferably 25°C to 40°C. Culturing is continued until a maximum of the desired product has formed. This target is usually reached within 10 hours to 160 hours.

Methods for the determination of L-amino acids are known from the prior art. The analysis can thus be carried out,
30 for example, as described by Spackman et al. (Analytical Chemistry, 30, (1958), 1190) by anion exchange chromatography with subsequent ninhydrin derivatization, or it can be carried out by reversed phase HPLC, for example as described by Lindroth et al. (Analytical Chemistry
35 (1979) 51: 1167-1174).

The invention furthermore relates to a process for the fermentative preparation of an amino acid chosen from the group consisting of L-asparagine, L-threonine, L-serine, L-glutamate, L-glycine, L-alanine, L-cysteine, L-valine, L-methionine, L-isoleucine, L-leucine, L-tyrosine, L-phenylalanine, L-histidine, L-lysine, L-tryptophan and L-arginine, in particular L-lysine, using coryneform bacteria which in particular already produce one or more of the amino acids mentioned.

10 The following microorganism was deposited on 23.01.2001 as a pure culture at the Deutsche Sammlung für Mikroorganismen und Zellkulturen (DSMZ = German Collection of Microorganisms and Cell Cultures, Braunschweig, Germany) in accordance with the Budapest Treaty:

15 • *Escherichia coli* strain Top10/pCR2.1citAint as DSM 13998.

The present invention is explained in more detail in the following with the aid of embodiment examples.

The isolation of plasmid DNA from *Escherichia coli* and all techniques of restriction, Klenow and alkaline phosphatase treatment were carried out by the method of Sambrook et al. (Molecular Cloning. A Laboratory Manual, 1989, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, USA). Methods for transformation of *Escherichia coli* are also described in this handbook.

The composition of the usual nutrient media, such as LB or TY medium, can also be found in the handbook by Sambrook et al.

ExamplesExample 1

Preparation of a genomic cosmid gene library from *C. glutamicum* ATCC 13032

- 5 Chromosomal DNA from *C. glutamicum* ATCC 13032 was isolated as described by Tauch et al. (1995, Plasmid 33:168-179) and partly cleaved with the restriction enzyme Sau3AI (Amersham Pharmacia, Freiburg, Germany, Product Description Sau3AI, Code no. 27-0913-02). The DNA fragments were
- 10 dephosphorylated with shrimp alkaline phosphatase (Roche Molecular Biochemicals, Mannheim, Germany, Product Description SAP, Code no. 1758250). The DNA of the cosmid vector SuperCos1 (Wahl et al. (1987), Proceedings of the National Academy of Sciences, USA 84:2160-2164), obtained
- 15 from Stratagene (La Jolla, USA, Product Description SuperCos1 Cosmid Vektor Kit, Code no. 251301) was cleaved with the restriction enzyme XbaI (Amersham Pharmacia, Freiburg, Germany, Product Description XbaI, Code no. 27-0948-02) and likewise dephosphorylated with shrimp alkaline
- 20 phosphatase.

- The cosmid DNA was then cleaved with the restriction enzyme BamHI (Amersham Pharmacia, Freiburg, Germany, Product Description BamHI, Code no. 27-0868-04). The cosmid DNA treated in this manner was mixed with the treated ATCC13032
- 25 DNA and the batch was treated with T4 DNA ligase (Amersham Pharmacia, Freiburg, Germany, Product Description T4-DNA-Ligase, Code no.27-0870-04). The ligation mixture was then packed in phages with the aid of Gigapack II XL Packing Extract (Stratagene, La Jolla, USA, Product Description
- 30 Gigapack II XL Packing Extract, Code no. 200217).

For infection of the *E. coli* strain NM554 (Raleigh et al. 1988, Nucleic Acid Res. 16:1563-1575) the cells were taken up in 10 mM MgSO₄ and mixed with an aliquot of the phage suspension. The infection and titering of the cosmid

library were carried out as described by Sambrook et al. (1989, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor), the cells being plated out on LB agar (Lennox, 1955, Virology, 1:190) + 100 µg/ml ampicillin. After 5 incubation overnight at 37°C, recombinant individual clones were selected.

Example 2

Isolation and sequencing of the citA gene

The cosmid DNA of an individual colony was isolated with 10 the Qiaprep Spin Miniprep Kit (Product No. 27106, Qiagen, Hilden, Germany) in accordance with the manufacturer's instructions and partly cleaved with the restriction enzyme Sau3AI (Amersham Pharmacia, Freiburg, Germany, Product Description Sau3AI, Product No. 27-0913-02). The DNA 15 fragments were dephosphorylated with shrimp alkaline phosphatase (Roche Molecular Biochemicals, Mannheim, Germany, Product Description SAP, Product No. 1758250). After separation by gel electrophoresis, the cosmid fragments in the size range of 1500 to 2000 bp were 20 isolated with the QiaExII Gel Extraction Kit (Product No. 20021, Qiagen, Hilden, Germany).

The DNA of the sequencing vector pZero-1, obtained from Invitrogen (Groningen, The Netherlands, Product Description Zero Background Cloning Kit, Product No. K2500-01) was 25 cleaved with the restriction enzyme BamHI (Amersham Pharmacia, Freiburg, Germany, Product Description BamHI, Product No. 27-0868-04). The ligation of the cosmid fragments in the sequencing vector pZero-1 was carried out as described by Sambrook et al. (1989, Molecular Cloning: A 30 Laboratory Manual, Cold Spring Harbor), the DNA mixture being incubated overnight with T4 ligase (Pharmacia Biotech, Freiburg, Germany). This ligation mixture was then electroporated (Tauch et al. 1994, FEMS Microbiol Letters, 123:343-7) into the E. coli strain DH5αMCR (Grant, 1990,

Proceedings of the National Academy of Sciences, U.S.A., 87:4645-4649) and plated out on LB agar (Lennox, 1955, Virology, 1:190) with 50 µg/ml zeocin.

The plasmid preparation of the recombinant clones was
5 carried out with Biorobot 9600 (Product No. 900200, Qiagen, Hilden, Germany). The sequencing was carried out by the dideoxy chain termination method of Sanger et al. (1977, Proceedings of the National Academies of Sciences, U.S.A., 74:5463-5467) with modifications according to Zimmermann et
10 al. (1990, Nucleic Acids Research, 18:1067). The "RR dRhodamin Terminator Cycle Sequencing Kit" from PE Applied Biosystems (Product No. 403044, Weiterstadt, Germany) was used. The separation by gel electrophoresis and analysis of the sequencing reaction were carried out in a "Rotiphoresis
15 NF Acrylamide/Bisacrylamide" Gel (29:1) (Product No. A124.1, Roth, Karlsruhe, Germany) with the "ABI Prism 377" sequencer from PE Applied Biosystems (Weiterstadt, Germany).

The raw sequence data obtained were then processed using
20 the Staden program package (1986, Nucleic Acids Research, 14:217-231) version 97-0. The individual sequences of the pZero1 derivatives were assembled to a continuous contig. The computer-assisted coding region analyses were prepared with the XNIP program (Staden, 1986, Nucleic Acids
25 Research, 14:217-231). Further analyses were carried out with the "BLAST search program" (Altschul et al., 1997, Nucleic Acids Research, 25:3389-3402) against the non-redundant databank of the "National Center for Biotechnology Information" (NCBI, Bethesda, MD, USA).

30 The resulting nucleotide sequence is shown in SEQ ID No. 1. Analysis of the nucleotide sequence showed an open reading frame of 1653 bp, which was called the citA gene. The citA gene codes for a polypeptide of 551 amino acids.

Example 3

Preparation of an integration vector for integration
mutagenesis of the citA gene

From the strain ATCC 13032, chromosomal DNA was isolated by
5 the method of Eikmanns et al. (Microbiology 140: 1817 -
1828 (1994)). On the basis of the sequence of the citA gene
known for *C. glutamicum* from example 2, the following
oligonucleotides were chosen for the polymerase chain
reaction (see SEQ ID No. 4 and SEQ ID No. 5):

10 citA-int1:

5` TTC CAG TCG GTG AGG TCA GT 3`

citA-int2:

5` GTA CGA TCG CGG ATG GTT AC 3`

The primers shown were synthesized by MWG Biotech
15 (Ebersberg, Germany) and the PCR reaction was carried out
by the standard PCR method of Innis et al. (PCR protocols.
A guide to methods and applications, 1990, Academic Press)
with the Taq-polymerase from Boehringer Mannheim (Germany,
Product Description Taq DNA polymerase, Product No. 1 146
20 165). With the aid of the polymerase chain reaction, the
primers allow amplification of an internal fragment of the
citA gene 480 bp in size. The product amplified in this way
was tested electrophoretically in a 0.8% agarose gel.

The amplified DNA fragment (see SEQ ID No. 3) was ligated
25 with the TOPO TA Cloning Kit from Invitrogen Corporation
(Carlsbad, CA, USA; Catalogue Number K4500-01) in the
vector pCR2.1-TOPO (Mead et al. (1991) Bio/Technology
9:657-663).

The *E. coli* strain TOP10 was then electroporated with the
30 ligation batch (Hanahan, In: DNA cloning. A practical
approach. Vol.I. IRL-Press, Oxford, Washington DC, USA,
1985). Selection for plasmid-carrying cells was made by
plating out the transformation batch on LB agar (Sambrook

et al., Molecular cloning: a laboratory manual. 2nd Ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1989), which had been supplemented with 50 mg/l kanamycin. Plasmid DNA was isolated from a transformant
5 with the aid of the QIAprep Spin Miniprep Kit from Qiagen and checked by restriction with the restriction enzyme EcoRI and subsequent agarose gel electrophoresis (0.8%). The plasmid was called pCR2.1citAint and is shown in figure 1.

10 The following microorganism was deposited as a pure culture on 23.01.2001 at the Deutsche Sammlung für Mikroorganismen und Zellkulturen (DSMZ = German Collection of Microorganisms and Cell Cultures, Braunschweig, Germany) in accordance with the Budapest Treaty:

15 • *Escherichia coli* Top10/pCR2.1citAint as DSM 13998

Example 4

Integration mutagenesis of the *citA* gene in the strains DSM 5715 and FERM-BP 1763

The vector pCR2.1citAint mentioned in example 3 was
20 electroporated by the electroporation method of Tauch et al. (FEMS Microbiological Letters, 123:343-347 (1994)) into the strains *Corynebacterium glutamicum* DSM 5715 and *Brevibacterium lactofermentum* FERM-BP 1763. The strain DSM 5715 is an AEC-resistant lysine producer (EP-B-0435132),
25 and the strain FERM-BP 1763 is a mycophenolic acid-resistant valine producer (US-A-5,188,948). The vector pCR2.1citAint cannot replicate independently in DSM5715 and FERM-BP 1763 and is retained in the cell only if it has integrated into the chromosome of DSM 5715 or FERM-BP 1763.
30 Selection of clones with pCR2.1citAint integrated into the chromosome was carried out by plating out the electroporation batch on LB agar (Sambrook et al., Molecular cloning: a laboratory manual. 2nd Ed. Cold Spring

Harbor Laboratory Press, Cold Spring Harbor, N.Y.), which had been supplemented with 15 mg/l kanamycin.

For detection of the integration, the citAint fragment was labelled with the Dig hybridization kit from Boehringer by the method of "The DIG System Users Guide for Filter Hybridization" of Boehringer Mannheim GmbH (Mannheim, Germany, 1993). Chromosomal DNA of in each case a potential integrant was isolated by the method of Eikmanns et al. (Microbiology 140: 1817 - 1828 (1994)) and in each case cleaved with the restriction enzymes EcoRI, BamHI and HindIII. The fragments formed were separated by means of agarose gel electrophoresis and hybridized at 68°C with the Dig hybridization kit from Boehringer. The plasmid pCR2.1citAint mentioned in example 3 had been inserted into the chromosome of DSM5715 and into the chromosome of FERM-BP 1763 within the chromosomal citA gene. The strains were called DSM5715::pCR2.1citAint and FERM-BP 1763::pCR2.1citAint.

Example 5

20 Preparation of lysine

The C.glutamicum strain DSM5715::pCR2.1citAint obtained in example 4 was cultured in a nutrient medium suitable for the production of lysine and the lysine content in the culture supernatant was determined.

25 For this, the strain was first incubated on an agar plate with the corresponding antibiotic (brain-heart agar with kanamycin (25 mg/l)) for 24 hours at 33°C. Starting from this agar plate culture, a preculture was seeded (10 ml medium in a 100 ml conical flask). The complete
30 medium CgIII was used as the medium for the preculture.

Medium Cg III

NaCl	2.5 g/l
Bacto-Peptone	10 g/l
Bacto-Yeast extract	10 g/l
Glucose (autoclaved separately)	2% (w/v)

The pH is brought to pH 7.4

Kanamycin (25 mg/l) was added to this. The preculture was incubated for 16 hours at 33°C at 240 rpm on a shaking machine. A main culture was seeded from this preculture
5 such that the initial OD (660 nm) of the main culture is 0.1 OD. Medium MM was used for the main culture.

Medium MM

CSL (corn steep liquor)	5 g/l
MOPS (morpholinopropanesulfonic acid)	20 g/l
Glucose (autoclaved separately)	50g/l
Salts:	
(NH ₄) ₂ SO ₄	25 g/l
KH ₂ PO ₄	0.1 g/l
MgSO ₄ * 7 H ₂ O	1.0 g/l
CaCl ₂ * 2 H ₂ O	10 mg/l
FeSO ₄ * 7 H ₂ O	10 mg/l
MnSO ₄ * H ₂ O	5.0mg/l
Biotin (sterile-filtered)	0.3 mg/l
Thiamine * HCl (sterile-filtered)	0.2 mg/l

Leucine (sterile-filtered) 0.1 g/l

CaCO₃ 25 g/l

The CSL, MOPS and the salt solution were brought to pH 7 with aqueous ammonia and autoclaved. The sterile substrate and vitamin solutions were then added, and the CaCO₃ autoclaved in the dry state was added.

5 Culturing was carried out in a 10 ml volume in a 100 ml conical flask with baffles. Kanamycin (25 mg/l) was added. Culturing was carried out at 33°C and 80% atmospheric humidity.

After 72 hours, the OD was determined at a measurement
10 wavelength of 660 nm with a Biomek 1000 (Beckmann Instruments GmbH, Munich). The amount of lysine formed was determined with an amino acid analyzer from Eppendorf-BioTronik (Hamburg, Germany) by ion exchange chromatography and post-column derivatization with ninhydrin detection.

15 The result of the experiment is shown in table 1.

Table 1

Strain	OD(660)	Lysine HCl g/l
DSM5715	7.5	13.3
DSM5715::pCR2.1citAint	7.4	14.4

Example 6

Preparation of valine

20 The B. lactofermentum strain FERM-BP 1763::pCR2.1citAint obtained in example 4 was cultured in a nutrient medium

suitable for the production of valine and the valine content in the culture supernatant was determined.

For this, the strain was first incubated on an agar plate with the corresponding antibiotic (brain-heart agar with
5 kanamycin (25 mg/l)) for 24 hours at 33°C. Starting from this agar plate culture, a preculture was seeded (10 ml medium in a 100 ml conical flask). The complete medium CgIII was used as the medium for the preculture. Kanamycin (25 mg/l) was added to this. The preculture was incubated
10 for 16 hours at 33°C at 240 rpm on a shaking machine. A main culture was seeded from this preculture such that the initial OD (660nm) of the main culture was 0.1 OD. Medium MM was used for the main culture.

Medium MM

CSL (corn steep liquor)	5 g/l
MOPS (morpholinopropanesulfonic acid)	20 g/l
Glucose (autoclaved separately)	50g/l
Salts:	
(NH ₄) ₂ SO ₄	25 g/l
KH ₂ PO ₄	0.1 g/l
MgSO ₄ * 7 H ₂ O	1.0 g/l
CaCl ₂ * 2 H ₂ O	10 mg/l
FeSO ₄ * 7 H ₂ O	10 mg/l
MnSO ₄ * H ₂ O	5.0mg/l
Isoleucine (sterile-filtered)	0.1 g/l
Methionine (sterile-filtered)	0.1 g/l

Thiamine * HCl (sterile-filtered)	0.2 mg/l
Leucine (sterile-filtered)	0.1 g/l
CaCO ₃	25 g/l

The CSL (corn steep liquor), MOPS (morpholinopropanesulfonic acid) and the salt solution were brought to pH 7 with aqueous ammonia and autoclaved. The sterile substrate and vitamin solutions were then added, and the CaCO₃ autoclaved in the dry state was added.

Culturing was carried out in a 10 ml volume in a 100 ml conical flask with baffles. Kanamycin (25 mg/l) was added. Culturing was carried out at 33°C and 80% atmospheric humidity.

After 48 hours, the OD was determined at a measurement wavelength of 660 nm with a Biomek 1000 (Beckmann Instruments GmbH, Munich). The amount of valine formed was determined with an amino acid analyzer from Eppendorf-BioTronik (Hamburg, Germany) by ion exchange chromatography and post-column derivatization with ninhydrin detection.

The result of the experiment is shown in table 2.

Table 2

Strain	OD(660)	Valine HCl g/l
FERM-BP 1763	12.1	7.5
FERM-BP 1763::pCR2.1citAint	13.5	10.8

SEQUENCE PROTOCOL

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5 <120> New nucleotide sequences which code for the citA gene

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gacttttctg cattatgatc agaattgttg gcctgggact tcgcttcacg ctctgctgat 180

aatcgccccc gggggtagac atg tct gtt ggt gga tcc gac tgg aaa aac ttc 233

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10

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Lys Glu Val Asp Ile Ile Arg Phe Ala Thr Arg Ile Leu Val Ile Gln

15

20

25

40

gtg gct acc gtc gcg ttg gtg gta gct att tgc acc gga att ttc gca 329

Val Ala Thr Val Ala Leu Val Val Ala Ile Cys Thr Gly Ile Phe Ala

30

35

40

45

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Val Leu Met Met Asp Gln Met Lys Thr Glu Ala Glu His Thr Ala Leu

45

50

55

tcc atc gga cgt tcg gtg gca tcc aac ccg cag atc cgc gag gaa gta 425

50

Ser Ile Gly Arg Ser Val Ala Ser Asn Pro Gln Ile Arg Glu Glu Val

60

65

70

75

gcg ctt gat act caa aca gga gca aac cca tcg gcc gaa gaa tta gcc 473

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80

85

90

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95

100

105

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	Ile Gly Leu Lys Leu Cys Arg Ala Leu Ala Arg Ser His Gly Gly Asp		
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The following figure is attached:

Figure 1: Map of the plasmid pCR2.1citAint.

The abbreviations and designations used have the following meaning.

KmR:	Kanamycin resistance gene
EcoRI:	Cleavage site of the restriction enzyme EcoRI
HindIII:	Cleavage site of the restriction enzyme HindIII
BamHI:	Cleavage site of the restriction enzyme BamHI
citAint:	Internal fragment of the citA gene
ColE1:	Replication origin of the plasmid ColE1

5

10

15

Patent claims

1. An isolated polynucleotide from coryneform bacteria, comprising a polynucleotide sequence which codes for the citA gene, chosen from the group consisting of
 - 5 a) polynucleotide which is identical to the extent of at least 70 % to a polynucleotide which codes for a polypeptide which comprises the amino acid sequence of SEQ ID No. 2,
 - 10 b) polynucleotide which codes for a polypeptide which comprises an amino acid sequence which is identical to the extent of at least 70% to the amino acid sequence of SEQ ID No. 2,
 - c) polynucleotide which is complementary to the polynucleotides of a) or b), and
 - 15 d) polynucleotide comprising at least 15 successive nucleotides of the polynucleotide sequence of a), b) or c),the polypeptide preferably having the activity of the sensor kinase CitA.
- 20 2. A polynucleotide as claimed in claim 1, wherein the polynucleotide is a preferably recombinant DNA which is capable of replication in coryneform bacteria.
3. A polynucleotide as claimed in claim 1, wherein the polynucleotide is an RNA.
- 25 4. A polynucleotide as claimed in claim 2, comprising the nucleic acid sequence as shown in SEQ ID No. 1.
5. A DNA as claimed in claim 2 which is capable of replication, comprising
 - 30 (i) the nucleotide sequence shown in SEQ ID no. 1, or

- (ii) at least one sequence which corresponds to sequence (i) within the range of the degeneration of the genetic code, or
 - (iii) at least one sequence which hybridizes with the sequences complementary to sequences (i) or (ii), and optionally
 - (iv) sense mutations of neutral function in (i).
6. A process as claimed in claim 5,
w h e r e i n
the hybridization is carried out under a stringency corresponding to at most 2x SSC.
7. A polynucleotide sequence as claimed in claim 2, which codes for a polypeptide which comprises the amino acid sequence shown in SEQ ID No. 2.
8. A vector pCR2.1citAint, which
- 8.1. carries an internal fragment of the citA gene 480 bp in size, shown in SEQ ID. No. 3,
 - 8.2 the restriction map of which is reproduced in figure 1, and
 - 8.3 which is deposited in the E. coli strain Top10/pCR2.1citAint at the Deutsche Sammlung für Mikroorganismen und Zellenkulturen [German Collection of Microorganisms and Cell Cultures] under no. DSM 13998.
9. An internal fragment of the citA gene with a length of 480 bp, shown in SEQ ID No. 3.
10. A coryneform bacterium, in which the citA gene is attenuated, preferably eliminated, in particular by deletion.

11. A process for the preparation of L-amino acids, in particular L-lysine,
w h e r e i n
it comprises carrying out the following steps,
- 5 a) fermentation of the coryneform bacteria which produced the desired L-amino acid and in which at least the citA gene is attenuated,
- b) concentration of the desired product in the medium or in the cells of the bacteria and
- 10 c) isolation of the L-amino acid.
12. A process as claimed in claim 11,
w h e r e i n
bacteria in which further genes of the biosynthesis pathway of the desired L-amino acid are additionally
15 enhanced are employed.
13. A process as claimed in claim 11,
w h e r e i n
bacteria in which the metabolic pathways which reduce the formation of the desired L-amino acid are at least
20 partly eliminated are employed.
14. A process as claimed in claim 11,
w h e r e i n
expression of the polynucleotide(s) which codes (code) for the citA gene is reduced, in particular
25 eliminated.
15. A process as claimed in claim 11,
w h e r e i n
the regulatory (or catalytic) properties of the polypeptide for which the polynucleotide citA codes
30 are decreased.
16. A process as claimed in claim 11,
w h e r e i n
for the preparation of L-amino acids, in particular L-

lysine, bacteria in which at the same time one or more of the genes chosen from the group consisting of

16.1 the dapA gene which codes for dihydrodipicolinate synthase,

5 16.2 the gap gene which codes for glyceraldehyde 3-phosphate dehydrogenase

16.3 the zwf gene which codes for glucose 6-phosphate dehydrogenase,

10 16.4 the pyc gene which codes for pyruvate carboxylase,

16.5 the lysE gene which codes for lysine export,

16.6 the lysC gene which codes for a feed back resistant aspartate kinase,

16.7 the zwal gene which codes for the Zwal protein

15 is or are enhanced, preferably over-expressed, are fermented.

17. A process as claimed in claim 11,
w h e r e i n

20 at the same time one or more of the genes chosen from the group consisting of:

17.1 the pck gene which codes for phosphoenol pyruvate carboxykinase,

17.2 the pgi gene which codes for glucose 6-phosphate isomerase,

25 17.3 the poxB gene which codes for pyruvate oxidase

17.4 the zwa2 gene which codes for the Zwa2 protein

is or are attenuated.

18. A process as claimed in one or more of the preceding
claims,
w h e r e i n
microorganisms of the genus *Corynebacterium glutamicum*
5 are employed.
19. A process for discovering RNA, cDNA and DNA in order
to isolate nucleic acids or polynucleotides or genes
which code for sensor kinase CitA or have a high
similarity with the sequence of the citA gene,
10 w h e r e i n
it comprises employing the polynucleotides comprising
the sequences according to claims 1 to 4 as
hybridization probes.

New nucleotide sequences which code for the citA gene**Abstract**

The invention relates to isolated polynucleotides comprising a polynucleotide sequence chosen from the group
5 consisting of

- a) polynucleotide which is identical to the extent of at least 70 % to a polynucleotide which codes for a polypeptide which comprises the amino acid sequence of SEQ ID No. 2,
- 10 b) polynucleotide which codes for a polypeptide which comprises an amino acid sequence which is identical to the extent of at least 70 % to the amino acid sequence of SEQ ID No. 2,
- c) polynucleotide which is complementary to the
15 polynucleotides of a) or b), and
- d) polynucleotide comprising at least 15 successive nucleotides of the polynucleotide sequence of a), b) or c),

and a process for the fermentative preparation of L-amino
20 acids using coryneform bacteria in which at least the citA gene is present in attenuated form, and the use of polynucleotides which comprise the sequences according to the invention as hybridization probes.

Figure 1: Plasmid pCR2.1citAint

